

Infection Prevention and Control Guidelines for Anesthesia Care

Chapter XX: Equipment and Environmental Cleaning, Disinfection, and Sterilization

Introduction

The following guidelines outline essential practices for equipment and environmental cleaning, disinfection, and sterilization in healthcare settings. This information is not exhaustive; for comprehensive guidance, refer to the CDC *Guidelines for Disinfection and Sterilization in Healthcare Facilities*, as well as applicable federal, state, and local law and regulations, manufacturer recommendations, and facility policies and procedures.¹

The following are general considerations for equipment and environmental cleaning and should not substitute review and adherence to previous referenced resources:

- **Policy Development:** Facilities should establish infection control policies specific to disinfection and sterilization of anesthesia equipment and monitor compliance regularly.¹⁻⁶
- **Disinfectant Selection:** Use U.S. Environmental Protection Agency (EPA)-registered disinfectants according to manufacturer instructions regarding concentration, contact time, disposal, and safety.^{7,8}
- **Cleaning Protocols:** Anesthesia equipment should be adequately cleaned prior to disinfection and sterilization.^{3,5,6,8}
- **Staff Training:** Ensure staff are trained in proper cleaning and disinfection techniques, with regular competency assessments.
- **Environmental Practices:** Minimize personal equipment (e.g., stethoscopes) and belongings (e.g., jackets, backpacks, bags, purses, personal electronic devices) in the operating room and/or patient care areas to reduce contamination risks.⁹

Purpose

To provide guidance on disinfection, sterilization, and waste management practices in healthcare settings, with a focus on anesthesia-related equipment and procedures.

Audience

This resource is intended for Certified Registered Nurse Anesthetists (CRNAs), also known as nurse anesthesiologists or nurse anesthetists, other anesthesia providers, members of the interdisciplinary team, administrators involved in policy developed, and other interested stakeholders.

The Spaulding Disinfection and Sterilization Classification Scheme

The Spaulding scheme classifies disinfection and sterilization methods for medical equipment by the risk of infection involved.^{1,2,10,11} View the details of the classification scheme in Table 1.

Table 1. Spaulding Disinfection and Sterilization Classification Scheme.

Device Classification	Device Example(s)	Process	Recommendation
Critical <i>Devices that enter sterile tissue, the vascular system, or through which blood flows.</i>	Surgical instruments, cardiac and urinary catheters, implants, and ultrasound probes used in sterile body cavities ^{1,11}	Sterilization ^{1,3, 4,8,10-12}	<ul style="list-style-type: none">• Sterilize between each patient use.• Use sterilization methods that destroy all vegetative bacteria, nonlipid viruses and bacterial spores.• Ensure thorough cleaning before sterilization.• Medical devices can be sterilized using chemical or physical properties depending on degree of contact with the patient.• Rinse with sterile water after chemical sterilization.³• Chemical germicides should be used rationally and in accordance with manufacturer recommendations and facility policy.
Semi-critical <i>Contact mucous membranes or non-intact skin.</i>	Anesthesia and respiratory therapy equipment, breathing circuits, endotracheal tubes, endoscopes, fiberoptic scopes, Magill forceps,	High-level disinfection ^{1,3, 4,8,10-13}	<ul style="list-style-type: none">• Clean thoroughly before disinfection.• Use FDA-approved high-level disinfectants that destroy all vegetative bacteria, mycobacteria, fungi, and viruses.• Rinse with sterile water after chemical disinfection.• Dry equipment using forced air or hanging to air

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Device Classification	Device Example(s)	Process	Recommendation
	cystoscopes ^{1,11}		<p>dry to prevent recontamination</p> <ul style="list-style-type: none"> • Dry all equipment surfaces to prevent humidity from encouraging microorganism growth. • Follow manufacturer's instructions for disinfectant concentration and contact time. • Store in a clean, dry area to prevent recontamination. • Perform high-level disinfection between patient uses.
	Laryngoscope blades ^{1,11,13-23}		<ul style="list-style-type: none"> • Wrap laryngoscope blades individually. • If high-level disinfection is used, a closed plastic bag may be used for storage. If steam sterilized, a peel pack may be used for storage. • Partially remove the blade from the package, attach to light source without touching blade surface, and test. Manipulation of the blade onto the light source/handle can be tested without actually removing the blade from the bag or pack without touching the blade itself. • Following testing, insert the blade back into the package and return to a clean, dry area to prevent recontamination. • Apply this protocol to both reusable and disposable blades.
	Laryngoscope handles ^{1,11,13,14,18-21,23}		<ul style="list-style-type: none"> • At a minimum, use intermediate-level disinfection with EPA-registered hospital disinfectant to wipe the handle after use. • Ensure adequate contact time as specified by the disinfectant manufacturer. • After disinfection, store in a clean, dry area to

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Device Classification	Device Example(s)	Process	Recommendation
			<p>prevent recontamination.</p> <ul style="list-style-type: none"> • Apply this protocol to both reusable and disposable handles. • Follow facility-specific policies and procedures for handle reprocessing. Some facilities may opt for high-level disinfection or sterilization based on risk assessment.
Non-critical <i>Contact intact skin.</i>	Patient Care Items: Electronic devices, stethoscopes, blood pressure cuffs, arm board, nametags, pulse oximeter sensors, head straps, monitor cables, blood warmers, medication administration pumps, carts, beds and monitors. ^{1,11}	Intermediate or low-level disinfection ^{1,11,12,24}	<ul style="list-style-type: none"> • Clean and disinfect between patients and when visibly soiled in accordance with manufacturer recommendations and facility policy. <ul style="list-style-type: none"> ◦ Low and intermediate-level disinfection differs by disinfectant type, concentration, and exposure to pathogen. • Use EPA-registered hospital disinfectants appropriate for the item. • Stethoscopes may be cleaned with soap and water, then disinfected with 70% isopropyl alcohol. • Use protective covering for non-critical surfaces that are difficult to clean (e.g., keyboard covers). • Hydrogen peroxide gas decontamination is an effective sterilization method for reusable items that are difficult to clean.
	Environmental Surfaces: Bed rails, food utensils, bedside furniture, computer keyboards, floors, mobile devices. ^{1,11}	Low-level disinfection (unless otherwise noted) ^{1,11,12}	<ul style="list-style-type: none"> • Clean and disinfect between patients and when visibly soiled in accordance with manufacturer recommendations and facility policy. • Use EPA-registered hospital disinfectants appropriate for the item. • Use protective covering for non-critical surfaces

Device Classification	Device Example(s)	Process	Recommendation
			that are difficult to clean (e.g., keyboard covers).

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46 **Single-Use Devices and Reprocessed Disposable Equipment**

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- A single-use device (SUD) is a medical device intended for use on one patient during a
 - 49 single procedure. It is not designed or validated for reuse.^{25,26} Studies have linked
 - 50 outbreaks of infection to improperly reprocessed medical devices.^{1,27-30}
 - 51 • Reuse of single-use devices may expose healthcare providers and facilities to additional
 - 52 liability.³¹
 - 53 • The reprocessing of SUDs is regulated by the FDA. Only FDA-registered third-party
 - 54 reprocessors or healthcare facilities that comply with FDA regulations may reprocess
 - 55 SUDs.^{25,26} Refer to the FDA for guidance and information on reprocessed single-use
 - 56 devices.^{1,25,26}

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58 To mitigate the incidence of outbreaks, it is recommended that healthcare facilities:

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- 60 • Develop and implement policies to verify the cleanliness and functionality of reprocessed
- 61 disposable equipment prior to use.^{25,26}
- 62 • Disassemble, clean, dry, reassemble, repackage, and disinfect or sterilize reprocessed,
- 63 disposable equipment prior to use as appropriate.^{25,26}
- 64 • Train staff on the proper handling and use of reprocessed SUDs.

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66 **The Anesthesia Machine and Breathing System**

67 Although there is no direct contact between the anesthesia workspace, anesthesia machine

68 controls and the patient, microorganisms can be transferred between the machine and patient

69 by the healthcare provider.^{23,32-36} Refer to federal, state or local law and regulations and facility

70 policies as well as specific manufacturer instructions for guidance concerning:^{3,8}

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- 72 • Cleaning and disinfecting the anesthesia machine.
- 73 • Pasteurizing or autoclaving of valves.

- Disassembling and disinfecting adjustable pressure-limiting valves.

Anesthesia Machine Surfaces and Carts^{3,8,23,32-34,36-39}

- Clean and disinfect anesthesia machine surfaces, knobs, and touchscreens between cases and at the end of each day using an EPA-registered hospital-grade disinfectant appropriate for the surface material.
- Implement measures to protect materials stored on the anesthesia machine from inadvertent contamination by airborne debris or fluids (e.g., blood).
- Regularly remove equipment from drawers, clean and disinfect the drawer interiors according to manufacturer's instructions
- Replace or clean the covering on the top of the anesthesia cart at the beginning of each case to maintain a clean work surface
- For small surfaces that are not visibly soiled, 70% isopropyl alcohol can be used for intermediate-level disinfection. However, for surfaces with visible contamination, use an EPA-registered disinfectant appropriate for the type of contamination.
- Clean carbon dioxide and soda lime absorbent canisters when changing the absorbent. Remove debris from screens and follow manufacturer's instructions for cleaning and maintenance.

Anesthesia Breathing System

Consult the manufacturer's user manual for specific cleaning recommendations for the breathing system.

Filters^{3,8,12,40-42}

Breathing system filters are single-use items evaluated by their bacterial filtration efficiency (BFE) and viral filtration efficiency (VFE).^{8,41} Bacterial filtration is generally more effective than viral filtration.^{3,8} Filters may prove problematic during spontaneous respiration due to increased resistance to air flow.³ While routine use of breathing system filters is not recommended due to inconclusive data on infection risk reduction, they should be used for patients with known respiratory infections (e.g., *Mycobacterium tuberculosis* infection).^{3,12}

- Consider placing high-efficiency filters on both inspiratory and expiratory limbs of the breathing circuit to protect both the patient and the anesthesia machine

- Filters may be placed between the endotracheal tube and the Y-piece.
- Use circuit filters and perform post-anesthesia machine disinfection after caring for patients with known pulmonary infections or trauma.

Carbon Dioxide Absorbers^{3,8,40,42}

- Follow manufacturer instructions for disassembly, cleaning, and sterilization of carbon dioxide absorbers.
- Clean canisters when changing the absorbent and remove debris from screens.
- Discard disposable plastic canisters after use.
- Periodically clean and disinfect bellows, unidirectional valves, and carbon dioxide absorbers.

Circuits^{1,3,8,41}

Anesthesia circuits may be single-patient use or multiple-patient use. For multiple-patient use circuits, place a new breathing system filter between the Y-piece and endotracheal tube after sterilization or high-level disinfection. Anesthesia professionals should pay close attention to anesthesia circuit product labeling.

- At a minimum, provide high-level disinfection for multiple-patient use breathing circuits.
 - If available, ultrasonic cleaning is effective.
- Disinfect the outer surface of the circuit between each use.
- Change end-tidal carbon dioxide tubing between patients.
- After anesthesia care for a patient with pulmonary infection or trauma, disinfect both internal and external components of the anesthesia machine's respiratory system.

Heat and Moisture Exchangers^{41,43-45}

- Heat and moisture exchangers alone are not effective in preventing microorganism transmission to the anesthesia breathing system.

Supraglottic Airway Devices^{21,46-52}

- If possible, use disposable single-use laryngeal mask airways (LMAs) due to the difficulty in completely removing protein deposits from reusable LMAs.
- If using reusable LMAs, rinse and soak in enzymatic detergent before autoclaving to remove occult blood.
 - Numerous studies have demonstrated that protein deposits are extremely difficult to eradicate completely from reusable LMAs.
- Consult manufacturer instructions for cleaning and sterilizing supraglottic airway devices.

Equipment Considerations for Special Patient Populations

Creutzfeldt-Jakob Disease^{3,12,53-62}

Creutzfeldt-Jakob Disease (CJD) is caused by prions, which are highly resistant to conventional sterilization methods. To properly disinfect equipment, consult the following recommendations:

- If lumbar puncture is being performed, limit the number of clinicians in the room to those who are essential.
- Attach a HEPA grade filter directly to the endotracheal tube.
- The gas sampling line leading to the respired gas analyzer should be connected only to the sampling port on the circuit side of the HEPA filter.
- Use disposable, single-use equipment whenever possible for patients with known or suspected CJD. Incinerate this equipment after use.
- For reusable equipment:
 - Destroy or quarantine laryngoscopes, supraglottic airway devices, and other instruments that have come into contact with high-risk tissues (e.g., spinal cord, eye).
 - Safely discard devices that are difficult or impossible to clean thoroughly.
- For instruments that must be reprocessed:
 - Clean thoroughly to remove visible contamination before sterilization.
 - Steam sterilize using one of the following methods:
 - Gravity displacement sterilizer: 132°C for 60 minutes
 - Prevacuum sterilizer: 134°C for 18 minutes
 - Alternative method for heat-sensitive instruments:

- Immerse in 1N sodium hydroxide for 1 hour at room temperature, rinse with water, then autoclave at 121°C for 30 minutes
- For noncritical items and environmental surfaces:
 - Decontaminate with 1N sodium hydroxide or sodium hypochlorite (i.e. bleach) at room temperature for 1 hour.
- Anesthesia machines used for CJD patients should undergo thorough decontamination, including disposal of the breathing circuit and carbon dioxide absorbent.
- Consult the CDC recommendations for best infection control practices when working with patients with CJD.

Tuberculosis^{3,63-67}

- Wear N95 respirators or higher-level respiratory protection.
- Place a HEPA filter between the breathing circuit Y-piece and patient's airway device.
- Perform high-level disinfection or sterilization on reusable equipment contacting patient's respiratory tract. Follow manufacturer's cleaning and sterilization instructions.
- Replace breathing circuit, reservoir bag, and carbon dioxide absorber after use.
- Routine anesthesia equipment culturing not recommended.

Environmental Surfaces

Healthcare facilities should establish and implement a comprehensive environmental cleaning and disinfection policy. This policy should specify:^{2,68-73}

- Frequency and level of disinfection (e.g., high-level, low-level)
- List of EPA-registered disinfectants approved for use in the facility
- Staff training on proper cleaning and disinfection techniques
- Procedures for monitoring compliance and performance improvement

Cleaning and disinfection practices:^{2,68,69,71-74}

- Procedures for monitoring compliance and performance improvement
- Clean and disinfect environmental surfaces to reduce transmission of HAIs from surfaces to providers and patients.

- Clean and disinfect anesthetizing locations and equipment surfaces (e.g., IV pumps, monitors, point-of-care devices, fluid warmers, forced air warmers) between patient cases and at the end of each day in accordance with facility policy.
- Follow manufacturer's recommendations for disinfectant use, contact time, and disposal.
- Place items that may be used during the next case on clean surfaces.

Product selection and use.^{68,69,71-74}

- Use EPA-registered hospital-grade disinfectants appropriate for the surface and suspected pathogens.
- Ensure proper contact time is maintained for effective disinfection.

Follow CDC recommendations for Standard Precautions and Transmission-Based Precautions.^{69,75}

Linens and Disposable Drapes⁷⁶⁻⁷⁸

- Handle linens and disposable drapes in a manner that minimizes the transfer of blood and microorganisms.
- Handle contaminated laundry as little as possible. Avoid unnecessary agitation of used linens to prevent aerosolization of pathogens.
- Place and transport the laundry in labeled or color-coded bags or containers.
- Do not sort or rinse contaminated laundry in patient care areas. Avoid body contact with soiled items; use appropriate PPE.
- Place and transport contaminated laundry in leak-resistant bags or containers.
- When using Standard Precautions, alternative labeling or color-coding is acceptable if it allows all personnel to recognize the containers as requiring compliance with precautions
- Store clean, laundered items in a clean, dry area to prevent contamination

Biohazardous Waste Management^{69,79-84}

Biohazardous waste refers to any item that is contaminated with infectious or potentially infectious materials. Sharps disposal is of particular concern due to the potential for injury when handling (e.g., needles, scalpel blades, drill bits, glass items).

- Dispose of all regulated waste in specified biohazard waste receptacles following federal, state, and local law and regulations.
- If a biohazardous waste container becomes contaminated, place the container inside of another biohazardous waste container.
- Consult relevant EPA and OSHA documents for specific guidance.

Single-Use Items

- Discard disposable single-use devices (e.g., breathing circuits, airway devices, orogastric tubes) in a biohazardous bag/container immediately after use.

Reprocessed Items

- Place items for reprocessing in a designated container immediately after use.
- Close containers before removing from the anesthetizing location.

Sharps Management

- Sharps include any device that may puncture skin (e.g., needles, syringes, scalpels, lancets, blades, glass).
- Use safety-engineered devices when possible.
- Do not bend or recap contaminated needles. If recapping is absolutely necessary, use a one-handed technique or mechanical device.
- Discard sharps immediately in a closeable, puncture-resistant, leak-proof container.

Drug Disposal

- Follow facility policy and applicable federal, state, and local law and regulations for disposal of partially remaining drugs in vials, ampules, syringes, and IV bags.
- Consider using EPA-registered pharmaceutical waste containers for certain medications.

The *Infection Control Guide for Certified Registered Nurse Anesthetists* was adopted by the AANA Board of Directors in 1992 and revised in 1993, 1997, November 2012. In February 2015, the AANA Board of Directors archived the guide and adopted the *Infection Prevention and Control Guidelines for Anesthesia Care*.

References

1. Rutala, WA, Weber, DJ, Healthcare Infection Control Practices Advisory Committee. Guideline for disinfection and sterilization in healthcare facilities. Atlanta, GA: Centers for Disease Control and Prevention; Jun 2024.
2. Guide to infection prevention for outpatient settings: Minimum expectations for safe care. Centers for Disease Control and Prevention. Accessed March 14, 2023.
www.cdc.gov/HAI/settings/outpatient/outpatient-care-guidelines.html
3. Dorsch J, Dorsch S. Cleaning and Sterilization. In: Brown B, ed. Understanding Anesthesia Equipment. Lippincott Williams and Wilkins; 2008:955-1000.
4. Rutala WA, Boyce JM, Weber DJ. Disinfection, sterilization and antisepsis: An overview. Am J Infect Control. Nov 2023;51(11s):A3-a12. doi:10.1016/j.ajic.2023.01.001
5. Kothekar AT, Kulkarni AP. Basic Principles of Disinfection and Sterilization in Intensive Care and Anesthesia and Their Applications during COVID-19 Pandemic. Indian J Crit Care Med. Nov 2020;24(11):1114-1124. doi:10.5005/jp-journals-10071-23562
6. Duan N, Gao W, Wang Q. Preparedness and disinfection of anesthetic equipment in COVID-19. J Clin Anesth. Nov 2020;66:109924. doi:10.1016/j.jclinane.2020.109924
7. Environmental Protection Agency. Selected EPA-Registered Disinfectants. Accessed Nov 15, 2024, <https://www.epa.gov/pesticide-registration/selected-epa-registered-disinfectants>
8. Juwarkar CS. Cleaning and sterilisation of anaesthetic equipment. Indian J Anaesth. Sep 2013;57(5):541-50. doi:10.4103/0019-5049.120152
9. Surgical Attire. In: Kyle E, ed. Guidelines for Perioperative Practice. Association of periOperative Registered Nurses; 2023:1087-1103.
10. Rutala WA, Weber DJ. Disinfection and Sterilization in Health Care Facilities: An Overview and Current Issues. Infect Dis Clin North Am. Sep 2016;30(3):609-37. doi:10.1016/j.idc.2016.04.002
11. Anderson M, Griffis C, Everson M, Reede L, Jeter L. Infection Control and Prevention. In: Elisha S, Heiner JS, Nagelhout JJ, eds. Nurse Anesthesia. 7th ed. Elsevier, Inc.; 2023:1372-1390:chap 61.
12. Sehulster L, Chinn RY. Guidelines for environmental infection control in health-care facilities. Recommendations of CDC and the Healthcare Infection Control Practices Advisory Committee (HICPAC). MMWR Recomm Rep. Jun 6 2003;52(RR-10):1-42.
13. Sherman JD, Hopf HW. Balancing Infection Control and Environmental Protection as a Matter of Patient Safety: The Case of Laryngoscope Handles. Anesth Analg. Aug 2018;127(2):576-579. doi:10.1213/ane.0000000000002759

14. Call TR, Auerbach FJ, Riddell SW, et al. Nosocomial contamination of laryngoscope handles: challenging current guidelines. *Anesth Analg*. Aug 2009;109(2):479-83.
doi:10.1213/ane.0b013e3181ac1080
15. Pino A, Lee JJ, Hashmi NK, Brucker A, Chow SC, Mahmood K. Prevention of contamination after endotracheal intubation using a dedicated sleeve. *J Thorac Dis*. Sep 28 2023;15(9):4717-4724.
doi:10.21037/jtd-22-1510
16. Van Wicklin SA. Contamination and Disinfection of Rigid Laryngoscopes: A Literature Review. *AORN J*. Jul 2019;110(1):49-59. doi:10.1002/aorn.12724
17. Gómez-Ríos M, Sastre JA, López T, Gaszyński T. Disinfection of Reusable Laryngoscopes: A Survey about the Clinical Practice in Spain. *Healthcare (Basel)*. Apr 13 2023;11(8)doi:10.3390/healthcare11081117
18. Chobin N. Protecting Disinfected Laryngoscope Blades in Storage. *Infection Control Today*. Accessed Nov 15, 2024, <https://www.infectioncontroltoday.com/view/protecting-disinfected-laryngoscope-blades-storage>
19. Negri de Sousa AC, Levy CE, Freitas MI. Laryngoscope blades and handles as sources of cross-infection: an integrative review. *J Hosp Infect*. Apr 2013;83(4):269-75.
doi:10.1016/j.jhin.2012.10.015
20. Weiers R. Protocol Example for Cleaning, High Level Disinfection, and Packaging of Laryngoscope Blades and Handles. General Hospital Supply Corporation. Accessed Nov 15, 2024, <https://www.ghscorp.net/blog/protocol-example-for-cleaning-high-level-disinfection-and-packaging-of-laryngoscope-blades-and-handles>
21. Ahmed SA, Fentie DY. Current practice of anesthetic equipment disinfection in the University of Gondar Comprehensive Specialized Hospital, 2020: a cross sectional study. *IJS Global Health*. 2021;4(3):e54. doi:10.1097/gh9.0000000000000054
22. Machan MD. Infection control practices of laryngoscope blades: a review of the literature. *Aana j*. Aug 2012;80(4):274-8.
23. Munoz-Price LS, Bowdle A, Johnston BL, et al. Infection prevention in the operating room anesthesia work area. *Infect Control Hosp Epidemiol*. Jan 2019;40(1):1-17. doi:10.1017/ice.2018.303
24. Petersson LP, Albrecht UV, Sedlacek L, Gemein S, Gebel J, Vonberg RP. Portable UV light as an alternative for decontamination. *Am J Infect Control*. Dec 2014;42(12):1334-6.
doi:10.1016/j.ajic.2014.08.012

25. Food and Drug Administration. Reprocessing Single-Use Medical Devices: Information for Health Care Facilities. Updated Aug 1, 2024. Accessed Nov 16, 2024, <https://www.fda.gov/medical-devices/products-and-medical-procedures/reprocessing-single-use-medical-devices-information-health-care-facilities>
26. Food and Drug Administration. Reprocessed Single-Use Devices: Frequently Asked Questions. Updated Aug 1, 2024. Accessed Nov 16, 2024, <https://www.fda.gov/medical-devices/reprocessing-single-use-medical-devices-information-health-care-facilities/reprocessed-single-use-devices-frequently-asked-questions>
27. Shuman EK, Chenoweth CE. Reuse of medical devices: implications for infection control. *Infect Dis Clin North Am*. Mar 2012;26(1):165-72. doi:10.1016/j.idc.2011.09.010
28. Garvey M. Medical Device-Associated Healthcare Infections: Sterilization and the Potential of Novel Biological Approaches to Ensure Patient Safety. *Int J Mol Sci*. Dec 22 2023;25(1)doi:10.3390/ijms25010201
29. Josephs-Spaulding J, Singh OV. Medical Device Sterilization and Reprocessing in the Era of Multidrug-Resistant (MDR) Bacteria: Issues and Regulatory Concepts. *Frontiers in Medical Technology*. Feb 10 2021;2doi:10.3389/fmedt.2020.587352
30. Mansur JM. Joint Commission International. Reuse of Single-Use Devices: Understanding Risks and Strategies for Decision-Making for Health Care Organizations. 2017;
31. Hearing Before the Subcommittee of Oversight and Investigations of the Committee on Commerce. Resuse of Single-Use Medical Devices. Serial No. 106–89. Feb 10, 2000
32. Baillie JK, Sultan P, Graveling E, Forrest C, Lafong C. Contamination of anaesthetic machines with pathogenic organisms. *Anaesthesia*. Dec 2007;62(12):1257-61. doi:10.1111/j.1365-2044.2007.05261.x
33. Loftus RW, Brown JR, Patel HM, et al. Transmission dynamics of gram-negative bacterial pathogens in the anesthesia work area. *Anesth Analg*. Apr 2015;120(4):819-26. doi:10.1213/ane.0000000000000626
34. Loftus RW, Koff MD, Burchman CC, et al. Transmission of pathogenic bacterial organisms in the anesthesia work area. *Anesthesiology*. Sep 2008;109(3):399-407. doi:10.1097/ALN.0b013e318182c855

35. Biddle CJ, George-Gay B, Prasanna P, Hill EM, Davis TC, Verhulst B. Assessing a Novel Method to Reduce Anesthesia Machine Contamination: A Prospective, Observational Trial. *Can J Infect Dis Med Microbiol.* 2018;2018:1905360. doi:10.1155/2018/1905360
36. Sanchez KL. Decreasing Anesthesia Workstation Contamination: An Educational Module. Nicole Wertheim College of Nursing Student Projects. Accessed Nov 16, 2024, <https://digitalcommons.fiu.edu/cnhs-studentprojects/150>
37. Loftus RW, Koff MD, Birnbach DJ. The dynamics and implications of bacterial transmission events arising from the anesthesia work area. *Anesth Analg.* Apr 2015;120(4):853-60. doi:10.1213/ane.0000000000000505
38. Anesthesia Patient Safety Foundation. FAQ on Anesthesia Machine Use, Protection, and Decontamination During the COVID-19 Pandemic. Updated July 27, 2023. Accessed Nov 16, 2024, <https://www.apsf.org/faq-on-anesthesia-machine-use-protection-and-decontamination-during-the-covid-19-pandemic/>
39. Schmidt E, Dexter F, Herrmann J, Godding JD, Hadder B, Loftus RW. Assessment of anesthesia machine redesign on cleaning of the anesthesia machine using surface disinfection wipes. *Am J Infect Control.* Jun 2020;48(6):675-681. doi:10.1016/j.ajic.2019.09.016
40. Spertini V, Borsoi L, Berger J, Blacky A, Dieb-Elschahawi M, Assadian O. Bacterial contamination of anesthesia machines' internal breathing-circuit-systems. *GMS Krankenhhyg Interdiszip.* 2011;6(1):Doc14. doi:10.3205/dgkh000171
41. Wilkes AR. Heat and moisture exchangers and breathing system filters: their use in anaesthesia and intensive care. Part 1 - history, principles and efficiency. *Anaesthesia.* Jan 2011;66(1):31-9. doi:10.1111/j.1365-2044.2010.06563.x
42. Macedo CE, Ferreira AM, Barcelos LDS, et al. Contamination of equipment and surfaces in the operating room anesthesia workspace: a cross-sectional study. *Sao Paulo Med J.* 2024;142(4):e2023177. doi:10.1590/1516-3180.2023.0177.R1.291123
43. Neft MW, Goodman JR, Hlavnicka JP, Veit BC. To reuse your circuit: the HME debate. *AANA J.* Oct 1999;67(5):433-9.
44. Auxiliadora-Martins M, Meneguetti MG, Nicolini EA, et al. Effect of heat and moisture exchangers on the prevention of ventilator-associated pneumonia in critically ill patients. *Braz J Med Biol Res.* Dec 2012;45(12):1295-300. doi:10.1590/s0100-879x2012007500161

45. Hallo-Carrasco A, Gruenbaum BF, Gruenbaum SE. Heat and Moisture Exchanger Occlusion Leading to Sudden Increased Airway Pressure: A Case Report Using ChatGPT as a Personal Writing Assistant. *Cureus*. Apr 2023;15(4):e37306. doi:10.7759/cureus.37306
46. Brimacombe J, Stone T, Keller C. Supplementary cleaning does not remove protein deposits from reusable laryngeal mask devices. *Can J Anaesth*. Mar 2004;51(3):254-7. doi:10.1007/BF03019106
47. Clery G, Brimacombe J, Stone T, Keller C, Curtis S. Routine cleaning and autoclaving does not remove protein deposits from reusable laryngeal mask devices. *Anesth Analg*. Oct 2003;97(4):1189-1191. doi:10.1213/01.ANE.0000080154.76349.5B
48. Miller DM, Youkhana I, Karunaratne WU, Pearce A. Presence of protein deposits on 'cleaned' reusable anaesthetic equipment. *Anaesthesia*. Nov 2001;56(11):1069-72. doi:10.1046/j.1365-2044.2001.02277.x
49. Coetzee GJ. Eliminating protein from reusable laryngeal mask airways. A study comparing routinely cleaned masks with three alternative cleaning methods. *Anaesthesia*. Apr 2003;58(4):346-53. doi:10.1046/j.1365-2044.2003.03084.x
50. Greenwood J, Green N, Power G. Protein contamination of the Laryngeal Mask Airway and its relationship to re-use. *Anaesth Intensive Care*. Jun 2006;34(3):343-6. doi:10.1177/0310057X0603400312
51. Bannon L, Brimacombe J, Nixon T, Keller C. Repeat autoclaving does not remove protein deposits from the classic laryngeal mask airway. *Eur J Anaesthesiol*. Jul 2005;22(7):515-7. doi:10.1017/s0265021505000888
52. Richards E, Brimacombe J, Laupau W, Keller C. Protein cross-contamination during batch cleaning and autoclaving of the ProSeal laryngeal mask airway. *Anaesthesia*. May 2006;61(5):431-3. doi:10.1111/j.1365-2044.2006.04550.x
53. Rutala WA, Weber DJ. Creutzfeldt-Jakob disease: recommendations for disinfection and sterilization. *Clin Infect Dis*. May 1 2001;32(9):1348-56. doi:10.1086/319997
54. Weber DJ, Rutala WA. Managing the risk of nosocomial transmission of prion diseases. *Curr Opin Infect Dis*. Aug 2002;15(4):421-5. doi:10.1097/00001432-200208000-00011
55. Centers for Disease Control and Prevention. Infection Control for CJD. Updated May 13, 2024. Accessed Nov 16, 2024, <https://www.cdc.gov/creutzfeldt-jakob/hcp/infection-control/index.html>
56. Brown P, Brandel JP, Sato T, et al. Iatrogenic Creutzfeldt-Jakob disease, final assessment. *Emerg Infect Dis*. Jun 2012;18(6):901-7. doi:10.3201/eid1806.120116

57. Washington State Department of Health. Human Prion Disease Infection Control Key Points. . Accessed Nov 16, 2024, <https://doh.wa.gov/sites/default/files/legacy/Documents/5100/420-162-PrionInfectionControl.pdf>
58. Belay ED, Blase J, Sehulster LM, Maddox RA, Schonberger LB. Management of neurosurgical instruments and patients exposed to Creutzfeldt-Jakob disease. *Infect Control Hosp Epidemiol*. Dec 2013;34(12):1272-80. doi:10.1086/673986
59. WHO Guidelines on Transmissible Spongiform Encephalopathies in relation to Biological and Pharmaceutical Products. Accessed Nov 16, 2024, <https://www.who.int/publications/m/item/who-guidelines-on-transmissible-spongiform-encephalopathies>
60. Commonwealth of Australia. Department of Health and Aged Care. Creutzfeldt–Jakob disease – Infection control guidelines. Updated Jan 16, 2013. Accessed Nov 16, 2024, <https://www.health.gov.au/resources/publications/creutzfeldt-jakob-disease-infection-control-guidelines>
61. UCSF Medical Center. Policies and Procedures for Patients with Suspected or Confirmed Human Prion Disease (e.g., Creutzfeldt-Jakob Disease [CJD]). Updated Mar, 2022. Accessed Nov 16, 2024, https://infectioncontrol.ucsfmedicalcenter.org/sites/g/files/tkssra4681/f/Sec%204.2%20Human%20Prion%20Policy_FINAL%2003-2022.pdf
62. Johnston L, Conly J. Creutzfeldt-Jakob disease and infection control. *Can J Infect Dis*. Nov 2001;12(6):332-6. doi:10.1155/2001/786564
63. Centers for Disease Control and Prevention. Tuberculosis Infection Control. Updated Dec 15, 2023. Accessed Nov 17, 2024, <https://www.cdc.gov/tb-healthcare-settings/hcp/infection-control/index.html>
64. Jensen PA, Lambert LA, Iademarco MF, Ridzon R. Guidelines for preventing the transmission of *Mycobacterium tuberculosis* in health-care settings, 2005. *MMWR Recomm Rep*. Dec 30 2005;54(Rr-17):1-141.
65. Krueger P, Klos C, Paulsen AW, et al. Cross-Contamination Via Anesthesia Equipment? *Anesthesia Patient Safety Institute*. 2009;24(1)
66. University of Texas Medical Branch. 01.21 – Tuberculosis (TB) Control Program. Updated Nov 20, 2023. Accessed Nov 17, 2024, https://www.utmb.edu/policies_and_procedures/Non-

IHOP/Healthcare_Epidemiology/01.21%20-

%20Tuberculosis%20(TB)%20Control%20Program.pdf

67. UNC Medical Center. Tuberculosis Control Plan. Updated Oct, 2022. Accessed Nov 17, 2024,
<https://spice.unc.edu/wp-content/uploads/2022/12/Tuberculosis-Control-Plan.pdf>

68. Rutala WA, Weber DJ. Best practices for disinfection of noncritical environmental surfaces and
equipment in health care facilities: A bundle approach. *Am J Infect Control*. Jun 2019;47s:A96-a105.
doi:10.1016/j.ajic.2019.01.014

69. Siegel JD, Rhinehart E, Jackson M, Chiarello L, Health Care Infection Control Practices Advisory C.
2007 Guideline for Isolation Precautions: Preventing Transmission of Infectious Agents in Health
Care Settings. *Am J Infect Control*. Dec 2007;35(10 Suppl 2):S65-164. doi:10.1016/j.ajic.2007.10.007

70. Centeleghe I, Norville P, Maillard JY, Hughes L. Infection prevention control in practice: a survey of
healthcare professionals' knowledge and experiences. *Infect Prev Pract*. Jun 2024;6(2):100357.
doi:10.1016/j.infpip.2024.100357

71. Link T. Guidelines in Practice: Environmental Cleaning. *Aorn j*. May 2021;113(5):487-499.
doi:10.1002/aorn.13376

72. Facciola A, Pellicano GF, Visalli G, et al. The role of the hospital environment in the healthcare-
associated infections: a general review of the literature. *Eur Rev Med Pharmacol Sci*. Feb
2019;23(3):1266-1278. doi:10.26355/eurrev_201902_17020

73. Dallolio L, Raggi A, Sanna T, et al. Surveillance of Environmental and Procedural Measures of
Infection Control in the Operating Theatre Setting. *Int J Environ Res Public Health*. Dec 28
2017;15(1)doi:10.3390/ijerph15010046

74. Weber DJ, Anderson D, Rutala WA. The role of the surface environment in healthcare-associated
infections. *Curr Opin Infect Dis*. Aug 2013;26(4):338-44. doi:10.1097/QCO.0b013e3283630f04

75. Centers for Disease Control and Prevention. Transmission-Based Precautions Accessed May 18,
2024, [https://www.cdc.gov/infection-control/hcp/basics/transmission-based-](https://www.cdc.gov/infection-control/hcp/basics/transmission-based-precautions.html)
[precautions.html](https://www.cdc.gov/infection-control/hcp/basics/transmission-based-precautions.html)

76. Centers for Disease Control and Prevention. Appendix D - Linen and laundry management. Updated
Mar 19, 2024. Accessed Nov 17, 2024, [https://www.cdc.gov/healthcare-associated-](https://www.cdc.gov/healthcare-associated-infections/hcp/cleaning-global/appendix-d.html)
[infections/hcp/cleaning-global/appendix-d.html](https://www.cdc.gov/healthcare-associated-infections/hcp/cleaning-global/appendix-d.html)

77. Centers for Disease Control and Prevention. G. Laundry and Bedding. Updated Jan 8, 2024. Accessed Nov 17, 2024, <https://www.cdc.gov/infection-control/hcp/environmental-control/laundry-bedding.html>
78. Association of Surgical Technologists. AST Standards of Practice for Surgical Drapes. 2008;
79. Occupational Safety and Health Administration 1910.1030 - Bloodborne Pathogens (2019).
80. Occupational Safety and Health Administration. Bloodborne Pathogens and Needlestick Prevention. Accessed May 18, 2024, <https://www.osha.gov/bloodborne-pathogens>
81. Occupational Safety and Health Administration. Bloodborne Pathogens and Needlestick Prevention General Guidance. Accessed Nov 17, 2024, <https://www.osha.gov/bloodborne-pathogens/general>
82. Denault D, Gardner H. OSHA Bloodborne Pathogen Standards. StatPearls. 2022.
83. Environmental Protection Agency. Medical Waste. Updated May 17, 2024. Accessed Nov 17, 2024, <https://www.epa.gov/rcra/medical-waste>
84. Environmental Protection Agency. Waste & Debris Fact Sheets. Accessed Nov 17, 2024, <https://iwaste.epa.gov/guidance/natural-disaster/fact-sheets/types-of-waste?id=biohazard-waste>